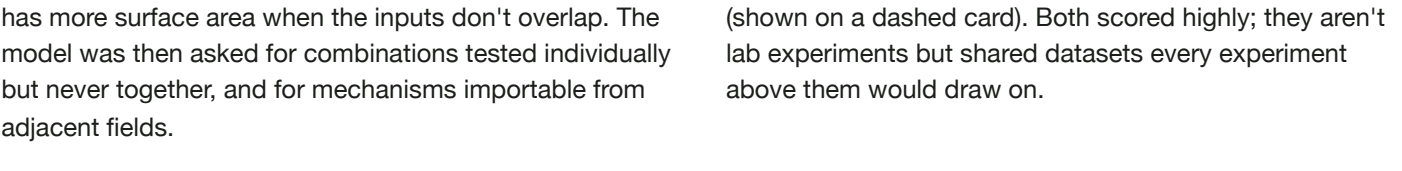


# Ten cross-disciplinary experiments for egg replacement & off-flavor reduction

A shortlist of benchtop-scale experiments generated by mining the seams *between* sub-literatures — enzyme chemistry, brewing, pharma formulation, cheese ripening — that individually study alt-protein problems but rarely cite one another.

**Corpus:** 80 peer-reviewed papers (40 off-flavor · 40 egg)  
**Method:** keyword-scored literature mine → LLM synthesis, cited to paper IDs  
**Selection:** top 5 experiments + 1 data program per field      **Status:** unbuilt — for specialist review

The premise is that the highest-leverage unexplored experiments in alternative-protein R&D are not the new mechanisms — they are **known mechanisms that have never been introduced to each other**. A phospholipase used for decades on egg yolk has never been pointed at mung-bean protein. Brewing solved aroma-loading of a protein matrix a century ago; plant-protein flavor science doesn't cite it. So rather than ask a model to invent chemistry, we built a small, tightly-filtered corpus and forced every proposed experiment to name the specific papers — and specific fields — it stitches together.



## THE GENERATIVE MOVE

The seed papers were hand-picked for *coverage of distinct intervention classes* (breeding, processing, enzymatic, fermentation, non-thermal, hybrid, precision fermentation) — deliberately *not* the top-scored papers, which all cluster on one well-studied intervention. Cross-paper synthesis has more surface area when the inputs don't overlap. The model was then asked for combinations tested individually but never together, and for mechanisms importable from adjacent fields.

## HOW TO READ THE RANKING

Each experiment carries a rough portfolio score — the product of five 1–5 judgments: *insight*, *probability of success*, *commercial value*, *breadth*, and *neglect*. Within each field the five benchtop experiments come first, ordered best-first — then one *data-infrastructure* program (shown on a dashed card). Both scored highly; they aren't lab experiments but shared datasets every experiment above them would draw on.

Where the ideas come from — the recurring pattern is a citation gap between two mature fields: phospholipase / egg-yolk chemistry × plant-protein design · brewing hop-stand aroma-loading × pea-protein flavor · cheese-rind ripening cultures × textured plant protein · pharma cyclodextrin/MIP capture × protein isolation · precision-fermented ovalbumin × plant-protein backbones · wine/tea tannin-aldehyde chemistry × protein slurries

## The ten experiments at a glance

| #                                                      | EXPERIMENT                                                                                  | CROSS-FIELD GRAFT            | EST. COST  | DIFFICULTY | P(SUCCESS) |
|--------------------------------------------------------|---------------------------------------------------------------------------------------------|------------------------------|------------|------------|------------|
| ● EGG REPLACEMENT — HARDEST UNSOLVED FUNCTIONS FIRST   |                                                                                             |                              |            |            |            |
| E1                                                     | H14 · <b>PLA2-hydrolyzed mung-bean liquid egg</b>                                           | enzyme chem → plant protein  | \$25–50k   | Moderate   | High       |
| E2                                                     | H15 · <b>PLA2 + transglutaminase cake-binder cascade</b>                                    | 2-enzyme cascade             | \$20–40k   | Moderate   | High       |
| E3                                                     | H16 · <b>2–5% ovalbumin spike on a plant backbone</b>                                       | precision ferm. × plant      | \$40–80k   | Moderate   | High*      |
| E4                                                     | H20 · <b>Hot-set + cold-set "boiled egg white" mimic</b>                                    | colloid gel cascade          | \$25–50k   | Moderate   | Moderate   |
| E5                                                     | H17 · <b>Aggregate-engineered mung foam for meringue</b>                                    | colloid sci → foam           | \$20–40k   | Moderate   | Mod–High   |
| E6                                                     | H21 · <b>Cross-protein egg-function benchmarking atlas</b><br><small>(DATA PROGRAM)</small> | shared measurement layer     | \$100–200k | Moderate   | Very high  |
| ● OFF-FLAVOR REDUCTION — HIGHEST-LEVERAGE LEVERS FIRST |                                                                                             |                              |            |            |            |
| O1                                                     | H1 · <b>Glycosidase knockdown for residual 1-octen-3-ol</b>                                 | glycoside pathway × breeding | \$50–100k  | Moderate   | Moderate   |
| O2                                                     | H2 · <b>Sequential RF + cold-plasma deactivation</b>                                        | two non-thermal levers       | \$25–75k   | Moderate   | High       |
| O3                                                     | H7 · <b>Cyclodextrin volatile-scavenging at isolation</b>                                   | pharma excipient → protein   | \$10–20k   | Simple     | High       |
| O4                                                     | H10 · <b>Glutaminase → alcalase → TGase enzyme cascade</b>                                  | 3-enzyme series              | \$30–60k   | Moderate   | Moderate   |
| O5                                                     | H8 · <b>Three-stage flavor-building fermentation</b>                                        | cheese/cured-meat cultures   | \$30–60k   | Moderate   | High       |
| O6                                                     | H4 · <b>Variety × harvest × storage GC-MS atlas</b><br><small>(DATA PROGRAM)</small>        | lot-selection dataset        | \$150–300k | Moderate   | High       |

\* mechanistically high; economics hinge on recombinant-ovalbumin price. "Novelty" throughout means *not found in this 80-paper corpus* — several ideas may exist in patents (excluded here) or industry trade secret.

## Egg replacement

Emulsification, structural gelling, and foam are three different egg jobs no single plant protein does well. These target the hardest ones — liquid egg, boiled-egg-white, meringue — with mechanisms borrowed from enzyme chemistry and colloid science.

1

EGG · H14 · FLAGSHIP EXPERIMENT

**PLA2-hydrolyzed mung-bean liquid egg with tailored emulsion architecture**

Treating a mung-bean protein + phospholipid mixture with phospholipase A2 reproduces the lyso-phospholipid + protein architecture that gives PLA2-treated egg yolk its emulsifying power — yielding a plant liquid-egg whose emulsion stability matches or beats hydrolyzed yolk.

COST \$25–50k

TIMELINE ~3 mo

DIFFICULTY Moderate

P(SUCCESS) High

NOVELTY High in corpus

INPUTS all food-grade, off-shelf

■ CROSS-DISCIPLINARY ORIGIN

egg\_017 (2024): PLA2 on egg yolk lifts protein solubility 27%→91% by disintegrating lipoprotein micelles.  
egg\_026: polysaccharide-conjugated mung protein emulsifies well. egg\_029: pH-shift + TGase is today's best plant liquid-egg. No one has pointed the yolk enzyme at mung protein.

■ THE EXPERIMENT

Mung isolate + x soy lecithin at 5:1 / 10:1 / 20:1 protein:lecithin, dosed with commercial food-grade PLA2 (e.g. Lecitase Ultra). Incubate; benchmark against native mung, PLA2-treated yolk, and commercial mayo.

■ MECHANISM

PLA2 cleaves the sn-2 fatty acid from phosphatidylcholine, yielding lyso-PC (a potent emulsifier) + free fatty acid. In yolk these reorganize the lipoprotein micelle; in mung protein + added soy lecithin the same chemistry should self-assemble emulsion-active structures.

■ READS OUT ON

emulsion stability index      droplet size (SLS)

freeze-thaw cycles      heat-set gel G'

KILL IF PC→lyso-PC conversion <30% at 2 h, droplet size >2x the yolk control, or ESI no better than the pH-shift/TGase baseline.

2

EGG · H15

**PLA2 + transglutaminase cascade as a single cake-binder ingredient**

A two-enzyme cascade — PLA2 to generate emulsifier in situ from added lecithin, then TGase to cross-link the protein network — reproduces *both* the emulsifying and structural-gelling roles of egg in one bakery ingredient.

COST \$20–40k

DIFFICULTY Moderate

P(SUCCESS) High

NOVELTY High in corpus

BUILDS ON E1 substrate stage

■ CROSS-DISCIPLINARY ORIGIN

egg\_017 (PLA2 emulsification) unified with egg\_005, egg\_006, egg\_029 (TGase functionality gains in mung / pea protein). Single-enzyme studies handle emulsification and gelling separately; the cascade does both in sequence.

■ THE EXPERIMENT

Apply PLA2 then TGase to mung isolate + lecithin slurry; bake into a fixed pound-cake formula against either enzyme alone, whole egg, and a commercial egg replacer.

■ MECHANISM

PLA2 first makes lyso-PC + FFA emulsifier and better-solubilizes the protein; TGase then cross-links Lys-Gln residues on the now-accessible network. Order matters — PLA2 doesn't consume TGase substrates, so emulsification precedes structure-building.

■ READS OUT ON

cake volume      crumb image analysis

texture profile (TPA)      7-day moisture retention

WIN BAR Matches whole-egg cake volume & crumb while beating the commercial replacer on staling.

3

EGG · H16

**A 2–5% precision-fermented ovalbumin spike on a plant-protein backbone**

Adding only 2–5% recombinant ovalbumin on top of a 95% plant-protein base buys egg-mimic cake structure at a fraction of a pure-recombinant cost — reframing ovalbumin as a functional cross-linker additive, not a wholesale egg replacer.

COST \$40–80k

DIFFICULTY Moderate

P(SUCCESS) High, price-sensitive

NOVELTY Moderate

NEEDS ovalbumin sample access

■ CROSS-DISCIPLINARY ORIGIN

egg\_013 (2025) names downstream-processing cost as the binding constraint on recombinant ovalbumin.  
egg\_035 shows ovalbumin's SH groups are the *structure-forming* element in egg-white cake. Spend the expensive protein only where it's uniquely needed — cross-linking — and let plant protein do the rest.

■ THE EXPERIMENT

Fixed chickpea or mung backbone + ovalbumin at 0/1/2/5/10% of total protein, baked against whole-egg control — run alongside a cost model parameterized on ovalbumin \$/kg.

■ MECHANISM

Ovalbumin's free thiols form disulfide cross-links during bake, supplying the structural backbone egg white contributes. A small spike delivers cross-linker density without paying for ovalbumin to also emulsify, where plant proteins are already adequate.

■ READS OUT ON

cake volume & crumb      sensory vs whole egg

cost / functional unit

PRIZE Egg-equivalent structure at ≤5% spike — a 10–50× out in required recombinant loading.

4

EGG · H20

**Hot-set + cold-set gel cascade for a plant "boiled egg white"**

Pairing a hot-set protein (potato or mung) with a cold-set gelling agent (κ-carrageenan / low-acyl gellan) plus TGase yields a plant boiled-egg-white that holds shape after both cooking and chilling — an unsolved, wide-open format.

COST \$25–50k

DIFFICULTY Moderate

P(SUCCESS) Moderate

NOVELTY Mod–High

WHITE SPACE standalone egg mimic

■ CROSS-DISCIPLINARY ORIGIN

egg\_020 (polysaccharide + plant-protein gels), egg\_021 (TGase-crosslinked plant-dairy gels) and egg\_031 (casein–soy TGase gels) each build networks — but none target the boiled-egg-white brief, which uniquely needs hot-set and cold-set/DM and the right bite at once.

■ THE EXPERIMENT

Factorial: protein (potato / mung / soy isolate) × polysaccharide (κ-carrageenan / gellan / mix) × TGase on/off. Cook 90 °C × 20 min, chill to 4 °C, profile texture.

■ MECHANISM

Plant protein supplies the thermal gel formed during cook; carrageenan + Ca<sup>2+</sup> supplies a cold-set network that survives chilling; TGase adds covalent cross-links robust across both temperature extremes.

■ READS OUT ON

TPA hardness / springiness      cohesiveness

water release      bite vs hard-boiled egg

TARGET A slice that survives salad handling with hard-boiled-egg-white bite.

5

EGG · H17

**Aggregate-engineered mung foam for a full meringue replacer**

The exceptional foaming of phase-separated mung-protein colloids is targeted by deliberate aggregate-forming pre-treatments — mild Maillard or controlled fermentation — to fully replace egg white in meringue, the canonical hardest foam test.

COST \$20–40k

DIFFICULTY Moderate

P(SUCCESS) Mod–High

NOVELTY High in corpus

■ CROSS-DISCIPLINARY ORIGIN

egg\_054 (2024): liquid-liquid phase-separated mung colloids reach 324% foamability with a 400-min half-life, far beyond mildly-purified mung. The colloid-science mechanism (aggregates thicken the film, block lamellar drainage) predicts that any aggregate-promoting treatment should push it further — a link the food-science meringue literature hasn't made.

■ THE EXPERIMENT

Four mung variants — baseline, colloid mix, LAB-fermented + conditioned (mild glucose, 65 °C × 6 h), LAB-fermented + heat-aggregated — compared on foam and baked meringue.

■ MECHANISM

Larger, denser aggregates in the thin lamella slow drainage. Mild Maillard cross-links raise protein-protein interaction; fermentation-driven aggregation does the same via pH-shift and bacterial-protease modification.

■ READS OUT ON

foam overrun      half-life / drainage curves

interfacial viscosity      meringue volume & crispness

TARGET A baked meringue matching egg-white structure and crisp set.

## DATA-INFRASTRUCTURE PROGRAM · SHARED MEASUREMENT LAYER, NOT A BENCHTOP EXPERIMENT

6

EGG · H21 · DATA PROGRAM

**Cross-protein × cross-function egg-replacement benchmarking atlas**

One normalized dataset measuring every candidate protein against every egg function at common conditions — so formulators *pick* the right protein for each role instead of guessing across papers that can't be compared.

COST \$100–200k

DIFFICULTY Moderate

P(SUCCESS) Very high (engineering)

NOVELTY The project, not the science

VALUE compounds over time

■ CROSS-DISCIPLINARY ORIGIN

egg\_001, egg\_018 and egg\_027 each benchmark proteins — but at different concentrations, methods and applications, so no two results line up. The data exists; it has never been put on one axis.

■ THE BUILD

One standardized protocol across ~20 commercial plant + dairy ingredients × 8 egg functions (emulsification, foaming, gelling, water- and oil-holding, binding, browning, opacity), released as a public database.

■ WHAT MAKES IT DIFFERENT

Not a hypothesis to falsify — a measurement substrate. It tests no mechanism; it removes the guesswork *under* every card above, telling you which protein to reach for before you run any of them.

■ DELIVERS

protein × function matrix

ESI · overrun · G' · MGC · WHC · OHC      open database

PAYOFF Ingredient selection becomes an evidence lookup instead of a guess — and the reference grows more valuable as rows are added.

## Off-flavor reduction

The beany / green / earthy signature of plant proteins is a mix of LOX-derived volatiles, non-LOX enzymatic sources, and formed compounds. These attack it at different points — precursor, process, isolation, and fermentation — with levers imported from breeding, brewing, pharma, and cheese-making.

1

OFF-FLAVOR · H1 · HIGHEST UPSIDE

**Glycosidase knockdown to eliminate residual 1-octen-3-ol**

Knocking down the soybean β-glucosidase that releases 1-octen-3-ol from its glycoside precursor eliminates the dominant mushroom / earthy off-note that *survives* LOX-null breeding — the residual gap every LOX-knockout program hits.

COST \$50–100k (inhibitor route)

DIFFICULTY Moderate

P(SUCCESS) Moderate

NOVELTY High in corpus

UPSIDE field-wide step change

■ CROSS-DISCIPLINARY ORIGIN

Three papers, never connected: off\_047 — 1-octen-3-ol is released from a glycoside *independent* of LOX; off\_007 — it still ranks top by odor-activity value *after* LOX knockout; off\_000 — residual off-flavor in triple-LOX-null soy traces to a heat-labile "LOX-like enzyme." Together they name a druggable glycosidase as the next target.

■ THE EXPERIMENT

(a) Screen varietal panels for naturally low β-glucosidase activity; (b) in parallel, add food-compatible β-glucosidase inhibitors at the soak step vs a heat-inactivated control; (c) quantify the full odor-active panel by HS-SPME-GC-MS.

■ MECHANISM

A β-glucosidase active during soak/maceration liberates volatile 1-octen-3-ol from a non-volatile glucoside. Anything that inactivates it before hydration — heat, an inhibitor, or a low-activity genotype — should suppress the note.

■ READS OUT ON

1-octen-3-ol / OAV      mushroom/earthy panel score

protein solubility / foaming

TARGET 1-octen-3-ol OAV <0.1× the LOX-null baseline without damaging protein function.

2

OFF-FLAVOR · H2

**Sequential radio-frequency + cold-plasma deactivation**

RF seed pretreatment (kills the precursor-making enzyme) followed by DBD reduction plasma on the wet slurry (oxidizes the volatiles that slip through) gives additive off-flavor reduction while staying below thermal protein-damage thresholds.

COST \$25–75k

DIFFICULTY Moderate

P(SUCCESS) High

NOVELTY High in corpus

RIGS pilot-scale available

■ CROSS-DISCIPLINARY ORIGIN

off\_018 (2026): RF at ~75 °C inactivates 80% of LOX with no yield loss. off\_021 (2026): DBD cold plasma at 60% O<sub>2</sub> degrades 1-octen-3-ol, 2-pentylfuran and others — compounds formed *after* extraction. Both are 2026; the obvious combination hasn't had time to publish.

■ THE EXPERIMENT

2x2 factorial (RF on/off × plasma on/off) on one yellow-pea source: dehumid → RF → wet fractionation → cold plasma at 60% O<sub>2</sub>, vs conventional control.

■ MECHANISM

The two act on different timescales (seed vs slurry) and different compound classes (precursors vs formed volatiles). Independent failure modes → multiplicative, not merely additive, reduction.

■ READS OUT ON

hexanal / nonanal      1-octen-3-ol / 2-pentylfuran

sensory triangle test      solubility & foaming

TARGET Combined arm beats either single treatment on volatiles with functionality intact.

3

OFF-FLAVOR · H7 · CHEAPEST, CLEANEST

**Cyclodextrin in-situ scavenging of pyrazines & aldehydes at isolation**

Adding β- or γ-cyclodextrin during pea-protein extraction traps methoxypyrazines and key aldehydes as inclusion complexes that partition off at ultrafiltration — stripping volatiles while leaving protein structure untouched.

COST \$10–20k

DIFFICULTY Simple

P(SUCCESS) High mechanistically

NOVELTY High in corpus

REGULATORY food-grade (E459/E468)

■ CROSS-DISCIPLINARY ORIGIN

off\_014 (2023) pins methoxypyrazines as major pea-aroma contributors — trace, polar, and stubbornly bound in protein hydrophobic pockets. Cyclodextrins are the standard pharma/cosmetic tool for exactly this capture problem, and are entirely absent from the plant-protein literature.

■ THE EXPERIMENT

β-CD and γ-CD at 1/2/5% w/v added post-extraction, pre-UF, vs untreated control. Profile volatiles in both retentate and permeate; confirm no structural change by SDS-PAGE.

■ MECHANISM

Cyclodextrins form 1:1 inclusion complexes with small hydrophobic guests. Pyrazines, hexanal, nonanal and 1-octen-3-ol all fit β/γ-CD cavities; CD-bound volatiles leave with the permeate at ultrafiltration, unchanged protein stays in the retentate.

■ READS OUT ON

retentate volatiles ↓      permeate volatiles ↑

CD residue (HPLC)      solubility / SDS-PAGE

TARGET Meaningful volatile drop at legal CD dose with an unchanged protein fingerprint.

4

OFF-FLAVOR · H10

**Glutaminase → alcalase → transglutaminase enzyme cascade**

Running three enzymes in series on dry-fractionated pea protein hits a trio usually treated as incompatible: lose beany volatiles, gain solubility, *and* recover gelling — because each enzyme fixes what the previous one leaves.

COST \$30–60k

DIFFICULTY Moderate

P(SUCCESS) Moderate

NOVELTY High in corpus

UNIFIES 3 separate process choices

■ CROSS-DISCIPLINARY ORIGIN

Each enzyme has a champion paper and a trade-off: glutaminase (off\_008) removes hexanal/2-pentylfuran but not function; alcalase (off\_024) cuts off-flavor but peptides lose gelling; TGase (egg\_005) rebuilds gelling. No one runs them as one ordered cascade.

■ THE EXPERIMENT

Single-variable scan on one dry-fractionated pea: each enzyme alone, all pairwise, and the full cascade — measuring flavor, sensory and function after every step.

■ MECHANISM

Glutaminase deamidates surface Gln/Asn (raises solubility, frees off-flavor binding sites); alcalase cleaves remaining hydrophobic peptides (more flavor release); TGase recrosslinks the network. Order is load-bearing — TGase substrate availability depends on the prior hydrolysis.

■ READS OUT ON

hexanal / 2-pentylfuran ↓      solubility & foaming ↑

gel hardness / G' held

PRIZE One ingredient that is simultaneously de-flavored, soluble, and gelling.

5

OFF-FLAVOR · H8

**Three-stage fermentation that builds flavor, not just removes it**

Adding a third, cured-food culture stage to an existing two-stage de-flavoring ferment simultaneously strips beany volatiles *and* adds meaty / umami aroma — flipping fermentation from cost-of-defect-removal into a flavor-creation asset.

COST \$30–60k

DIFFICULTY Moderate

P(SUCCESS) High chem · Mod sensory

NOVELTY Mod–High

■ CROSS-DISCIPLINARY ORIGIN

off\_003 (2025) runs two LAB/yogurt stages that reduce volatiles but add nothing; off\_034 shows fermentation + protease yields amino-acid-derived aromas; off\_037 names rapid-acidifying strains. The natural third stage — surface-ripened-cheese and dry-cured-ham cultures — is unrun on plant protein.

■ THE EXPERIMENT

Common 9% pea substrate; identical stages 1–2; four stage-3 candidates (B. linens, S. carnosus, D. hansenii, control) with added protease. Track volatile evolution by SIFT-MS at each stage.

■ MECHANISM

Stages 1–2 (L. plantarum → yogurt culture) drop pH and degrade volatiles. Stage 3 (Brevibacterium linens or Staphylococcus carnosus + neutral protease) generates Strecker aldehydes and short-chain acids — the same chemistry behind cheese-rind and cured-ham aroma.

■ READS OUT ON

stagewise volatile fingerprint

beany ↓ / umami · meaty ↑      free amino acids (HPLC)

TARGET Net-positive aroma profile with beany notes suppressed and savory notes built.

## DATA-INFRASTRUCTURE PROGRAM · LOT-SELECTION DATASET, NOT A BENCHTOP EXPERIMENT

6

OFF-FLAVOR · H4 · DATA PROGRAM

**Variety × harvest-year × storage → volatile-fingerprint atlas**

A public dataset mapping how variety, harvest year, location and storage drive pea-protein volatile load — so formulators *select* low-off-flavor lots rather than spend yield and function process-correcting them.

COST \$150–300k-yr-1

DIFFICULTY Moderate

P(SUCCESS) High (engineering)

NOVELTY High in corpus

VALUE compounds over time

■ CROSS-DISCIPLINARY ORIGIN

off\_040 (harvest year & location dominate LOX activity), off\_039 (storage reshapes volatiles over a year) and off\_001 (varietal fatty-acid profile → off-note load) together imply lot selection can outperform process intervention — yet no selection infrastructure exists.

■ THE BUILD

3–5 processor partners each contribute ~20 lots/year with metadata; standardized PPI extraction + HS-SPME-GC-MS fingerprint; open-data release; a regression predicting volatile load from metadata, validated on held-out lots.

■ WHAT MAKES IT DIFFERENT

Not a process to test — a selection layer. Where the five experiments above *correct* off-flavor, this lets you *avoid* it upstream at zero yield cost, and quantifies how much of the problem is lot choice versus processing.

■ DELIVERS

volatile fingerprint per lot      metadata → volatile model

open dataset

PAYOFF Predict hexanal / nonanal / 1-octen-3-ol on unseen lots (R<sup>2</sup>>0.7 is useful) — buy the good lots, skip the correction.

## READING & PROVENANCE

**Paper IDs** (e.g. egg\_017, off\_047) are stable references into the project corpus; each maps to a title, abstract and DOI in the underlying dataset, so any claim above traces back to primary literature. "Novelty (in corpus)" means the exact experiment was not found among these 80 papers — it does not rule out prior art in patents (deliberately excluded) or unpublished industry work; treat every card as a hypothesis to be checked, not a claim of first-invention. **Costs, timelines and success odds** are order-of-magnitude buckets for triage, not quotes. **The sixth item in each field** appears in a dashed card because it is a different kind of thing — a public dataset (a variety-x-harvest-x-storage GC-MS atlas; a cross-protein functional-benchmarking database) rather than a benchtop test. Both score highly and would de-risk every experiment above them, but they should be judged as engineering programs, not hypotheses to falsify.

Alt-Protein Paper Scout · hypothesis brief v1 · generated for specialist review · not a funding submission.